

# Efficiency Evaluation of Anti-corrosion Treatment of Carbon Steel by Extracts of Red Algae Collected from Mediterranean Coast

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**Abstract.** *The paper presents the assessment of inhibition effectiveness of biomolecules extracted from red seaweed against biocorrosion in the petroleum industry. The first objective of this study was to obtain extracts (A, B and C, prepared respectively from red algae species: Corallina elongata, Gymnogongrus crenulatus and Pterocladia capillacea) by ethanol extraction method. The infrared spectra of the three extracts confirmed the presence of amine derivatives molecules known by their anti-corrosion inhibiting powers. The second objective was based on the identification of physico-chemical characteristics of the extracts and thus revealing their inhibitory and / or bactericidal power in bacterial corrosion of carbon steel in injection water contaminated with sulfato-reducing bacteria. Biological test of all extracts gave a concentration upto 10 germs/mL in contaminated water by sulfato-reducing bacteria during 28 days of incubation at 37°C. Evolution in time of the open-circuit potential showed a longer incubation time for electrolyte with extracts, whereas the stabilization time was shorter. Current corrosion density, polarization resistance, charge transfer resistance and double layer capacity were determined by using linear polarizarion resistance technique and electrochemical impedance spectroscopy. The corrosion protection efficiency of extract obtained from Gymnogongrus crenulatus (extract B) reached a maximum protective capacity of 99.69% at 5 ppm in the injection water.*

**Keywords:** *biocorrosion, inhibition, red algae, corrosive sulfato-reducing bacteria, electrochemical techniques*

## 1. Introduction

The degradation of metals under the influence of microorganisms, called biocorrosion, has significant repercussions on the environment and on economy in various industrial fields, notably the oil industry [1]. The materials used in petroleum field in contact with a wet environment are liable to be corroded because of presence of microorganisms, mainly sulfato-reducing bacteria SRB, which are the main cause of anaerobic corrosion by producing hydrogen sulfide (H<sub>2</sub>S) corrosive metabolite [2]. Certainly, in order to maintain pressure oil reservoir, it is necessary to apply a system making it possible to compensate for oil's volume extracted, this being the injection water. The physicochemical composition and the nutrient richness of this water present an environment favorable to microorganism's proliferation, in particular corrosive bacteria that adhere strongly to metallic surfaces forming a biofilm.

Various treatments have been carried out to combat metallic structures degradation (corrosion) and to preserve petroleum installations with disadvantages of the use of inhibitors (using inhibitors containing toxic compounds such as chromium and nitrites) use as much as inhibitors of complicated corrosion [3-5].

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Algae are groups of globally distributed photosynthetic organisms, usually aquatic or alive in wetlands. According to their pigments and other morphological and physiological characteristics, several classes of algae can be distinguished: brown algae, green algae (*Chlorophyceae*) and red algae (*Rhodophyceae*) [6]. Red algae are present in fresh water but mostly are marine, they are found in particular on the tropical and subtropical coasts. Their colors diverge from blackish to multiple shades of red; these tints arise from the presence of red phycoerythrin protein or blue Pyocyanin that masks chlorophyll [7,8]. In this study, we used three algae harvested in the Zemmouri el Bahri and Dellys regions of Boumerdes on the eastern Algerian coast in Mediterranean waters. One is *Corallina elongata* (syn. *Corallina mediterranea* or *Ellisolandia elongata*) from *Corallinaceae* family, an autotrophic marine red alga which inhabits light rocks (photophile species) in calm water and is very tolerant to pollution (opportunistic species) [9]. The second alga, *Gymnogongrus crenulatus* from *Phyllophoraceae* family, is hung on rocks generally in shallow water [9]. The third alga is *Pterocladia capillacea* from *Pterocladaceae* family, an autotrophic red-black alga that develops in the superficial rock faults of the infralittoral stage in dark, calm, semi-slaughtered mediterranean and atlantic environments [10]. The main objective of this work is to study inhibitory effect of three natural products extracted from marine red algae on biocorrosion of carbon steel immersed in injection water contaminated by SRB as well as identification of protective films formed on carbon steel surface by several electrochemical techniques: open-circuit potential (OCP) in time, linear polarization resistance (LPR) technique and electrochemical impedance spectroscopy (EIS).

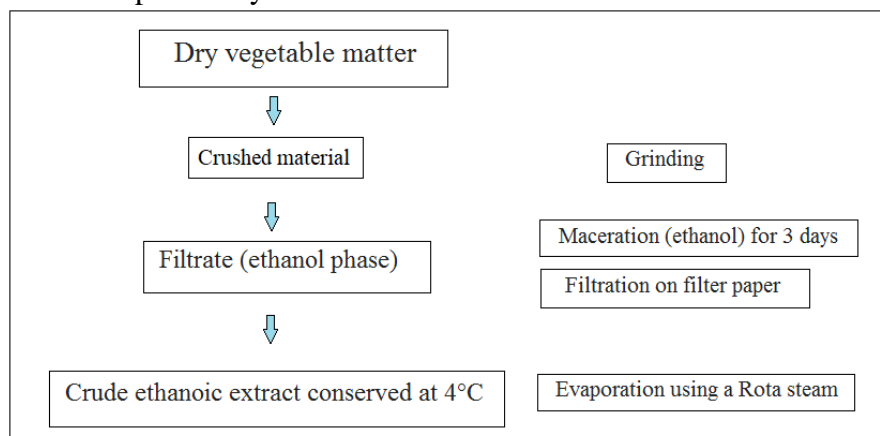
## 2. Materials and methods

### 2.1. Preparation of biological material

Red algae were collected in various regions; *Dellys* (Latitude 36.9158, Longitude 3.9131) and *Zemmouri El Bahri* coast (Latitude 36.8025, Longitude 3.56699) on December 2015 (Figure 1). Red algae collecting were carried out in places rich in algae, clinging to rocks located from 3 to 6m at the seashore and 20cm to one meter deep. 500g of the samples are dried in the open air at an ambient temperature for one month and then crushed using an electric grinder and stored in brown glass jars protected from light and moisture.

### 2.2. Characterization and extracting plant active material

The extraction of essential oils from marine red algae was carried out using an organic solvent, ethanol, therefore an "ethanol extraction" was made. The red algae collected were ground until powders were obtained. Ethanolic extract of algae was prepared by maceration under a ventilated hood, 90 g of red powder and 180 mL of ethanol are introduced into a 250 mL Erlenmeyer flask [11]. The flask was then closed to avoid evaporation of the solvent, and then continuously stirred at an average speed for three successive days (72 h), at ambient temperature. After stirring, the macerate was filtered through a funnel, where a compress was first placed, through a filter paper. Figure 1 summarizes the various extraction steps already mentioned.



**Figure 1.** Steps for obtaining ethanoic crude extract

After that, we determined the organoleptic and physicochemical properties (pH, density, viscosity, appearance, color and odor) of ethanolic extracts. Analysis by infrared spectroscopy made it possible to identify the active biomolecules constituting the three extracts using a spectrometer (Shimadzu 8900 type FTIR spectrometer).

The sample of injection water used for this study comes from the oilfield of *Rhoude El Baguel* located 70 km from east of *Hassi Messaoud* in the south of *Algeria*.

### 2.3. Efficiency tests of bionatural extracts A, B and C on growth of SRB by microbiological method

For evidencing the inhibitory effect, three representative algae extracts were prepared: **A** (from *Corallina ellongata*), **B** (from *Gymnogongrus crenulatus*) and **C** (from *Pterocladia capillacea*). The sample of injection water used for this study comes from the oilfield of *Rhoude El Baguel* located 70 km from east of Hassi Messaoud in the south of Algeria. Seven sets of five penicillin vials containing Postgate's SRB culture media and decimal dilutions of injection water were incubated for 48 h. After autoclaving, the first set of five was considered than control, and the remaining six series will serve as an inoculum for the efficacy trials of extracts extract s A, B and C obtained from the marine red algae. In order to evaluate the efficiency of these, two concentrations of the extracts A, B and C were injected into the penicillin vials prepared previously and then incubated at 37°C for 28 days. The results are expressed throughout the incubation period. During the visual examination of the vials, two cases occur; first case: Positive vial shift from staining to black (synthesis of H<sub>2</sub>S by SRB); ineffective extract and the second case: Negative bottle not blackening the extract is effective.

### 2.4. Inhibition efficiency test of bionatural extracts by weight loss method

To demonstrate the bactericidal efficacy of the three seaweed extracts, we carried out a test for the evaluation of the bacterial corrosion rate by weight loss method [12, 13]. After polishing carbon steel coupon's area of 1.44 cm<sup>2</sup>, the steel samples were immersed in injection water inoculated with SRB (10%). Bottles are placed in an incubator for 48 h and then 15 ppm of each extract was injected into the first three flasks and the fourth is considered as a control. The duration of exposure was 45 days. All weighings were performed with analytical balance (KERN). and were done using method of weight loss norm (RP 0775-99 - corrosion coupons installation and handling). This method is based on the calculation of the rate of corrosion with respect to the difference in weight of steel before and after its exposure in a corrosive medium during 45 days, based on the equation:

$$V_{\text{corr}} = (\Delta P / t \cdot \rho \cdot S)$$

where:  $V_{\text{corr}}$ : corrosion rate,  $\Delta P = P_f - P_i$  is weight loss,  $t$ : exposure time of the steel coupon (it was considered the number of 8760 h in a year),  $\rho$ : density of carbon steel (7.86 g/cm<sup>3</sup>),  $S$ : surface area of the corroded coupon.

The steel coupons are then observed using a scanning electron microscope (MEB-QUANTA 650) and a metallurgical microscope (Olympus PMG3).

### 2.5. Inhibition efficiency test of bionatural extracts by electrochemical method

The electrochemical equipment used in this study is composed of a potentiostat /galvanostat (Type EGEG Princeton Applied Research, model 273A), that makes it possible to impose and measure stable potentials and currents in cathodic and anodic domains (Electrometer; Type EGG, model 273A), Frequency analyzer of Solarton type (model SI 1255) for electrochemical impedance measurement. Nitrogen generator equipped with a pressure regulator with an inlet pressure of 300 bars and an outlet pressure adjustable between 1 and 12 bars, computer equipment connected to the potentiostat /galvanostat equipped with data acquisition, and processing software, making it possible to plot the various curves and to calculate the electrochemical parameters (Software 352 SoftCorrIII), for stationary methods and

Software Z plot, Z view, for electrochemical impedance spectroscopy and electrochemical cell, composed of Pyrex glass of volume 1000 mL, provided with five orifices for carrying the following accessories: A working electrode; A saturated calomel reference electrode (SCE); A glass gas bubbler; Two counter-electrodes in graphite 6 mm in diameter and 30 cm in length.

A volume of 700 mL of the injection was placed on an electrochemical cell made of Pyrex glass. The graphite electrodes were placed in the electrochemical cell, the other orifices were sealed. The cell was bubbled with nitrogen for 30 min in order to exclude all the air from the test solution and then autoclaved at 120°C for 20 min at 1 bar. In front of a Bunsen spout, inoculation of the electrochemical cell with 5% BSR of  $10^6$  germs/mL concentration was carried out, a reference electrode SCE (saturated calomel electrode) was introduced, and finally the carbon steel working electrode of API 5L Grade X60 type (Table 1) was immersed in the mixture after being polished and rinsed with acetone.

**Table 1.** Chemical composition of carbon steel (API 5L x60)

ELEMENT %	C	Si	Mn	Ni	Cr	P	S	Mo	Cu	Fe
API 5L X60	0.22	0.23	1.31	0.044	0.05	0.013	0.01	0.016	0.06	98.047

The bionatural extracts A, B and C at different concentrations then were treated the solution (5ppm, 10ppm, 15ppm, 20ppm, 30ppm, 50 ppm). The curves of the different techniques such as the open -circuit potential (OCP) vs. - time, linear polarization resistance curves (LPR) and electrochemical impedance spectra (EIS) were recorded. Open circuit potential OCP  $\pm$  30 mV for potential region, potential scan rate of 0.166 mV/s and EIS sinusoidal voltage of 10 mV applied with frequency domain from 100 KHz to 10 mHz were used.

Values of polarization resistance  $R_p$ , corrosion current density  $I_{Corr}$ , corrosion rate CR and inhibition efficiency E were calculated by using LPR technique. Values of the ohmic resistance of solution  $R_e$ , charge transfer resistance  $R_{ct}$ , double layer capacity  $C_{dl}$ , phase angle  $\theta$  and inhibition efficiency E were determined using EIS diagrams as Nyquist and Bode spectra. The inhibition efficiency of the extract was calculated from the charge transfer resistance values using the following equations:

$$E\% = \frac{R_{Po} - R_p}{R_{Po}} \cdot 100 \quad E\% = \frac{R_{cto} - R_{ct}}{R_{cto}} \cdot 100$$

where:  $R_{Po}$ ,  $R_p$  and  $R_{cto}$ ,  $R_{ct}$  are the charge transfer resistance in absence and in presence of inhibitor extract, respectively.

### 3. Results and discussions

#### 3.1. Characterization of algae extracts and electrolyte

The organoleptic and physicochemical analysis of the extract algae were done by examine

The appearance, viscosity, pH, solvent solubility and water solubility as per quality standards of European pharmacopoeia are shown in Table 2 and Figure 2.

**Table 2.** Organoleptic and physico-chemical characteristics of organic extracts a, b and c obtained from marine red algae

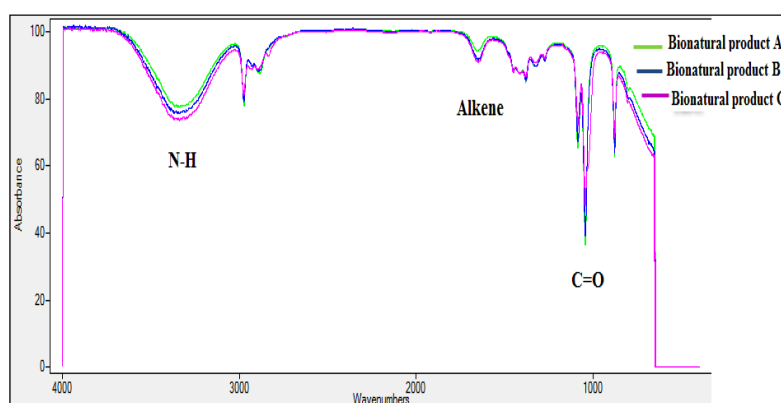
	Odor	Color	pH	Density g/cm <sup>3</sup> at 20°C	Viscosity cP	Solubility in		
						water	Alcohol	Hydrocarbon
Extract A	Characteristic of algae	green	6.9	0.831	1.72	+	+	-

Extract B	Characteristic of algae	dark green	6.3	0.851	1.91	+	+	-
Extract C	Characteristic of algae	dark green	6.1	0.866	2.18	+	+	-



**Figure 2.** Aspect of three algae extracts and solubility test results

The characterization of extracts by infrared spectroscopy was made in order to know their relative chemical compositions and thus revealing the structure of the bioactive molecules that compose them. Therefore, Figure 3 shows the adsorption bands of the extracts A, B and C based on marine red algae. The identification by infrared spectroscopy shows identical spectra for three extracts A, B and C. The spectrum exhibits an intense absorption band in the region  $1820\text{--}1660\text{ cm}^{-1}$  indicates the presence of carboxyl group  $\text{C}=\text{O}$ . We observed an absorption band of average intensity around  $3400$  and  $3300\text{ cm}^{-1}$  indicates presence of group  $\text{N-H}$ , confirms the presence of amine function in the extract. The spectrum exhibits a low intensity absorption band located at about  $1650\text{ cm}^{-1}$  indicates the presence of the alkene function. This is probably the spectrum of the trimethylamine molecule, which has the formula  $(\text{C}_3\text{H}_9\text{N})$ . Amines are very often used for the protection of parts in natural aqueous media they are considered as functional groups very effective in corrosion of ferrous metals these organic compounds capable of functioning as corrosion inhibitors contain an active center N, S, O, P capable of exchanging electrons with metal [14].



**Figure 3.** Infrared spectrum of three bionaturals extracts A, B and C

Injection water was obtained from the of *Rhoude El Baguel* oilfield. *In situ*, the injected water temperature is between  $19$  and  $20^\circ\text{C}$  and  $\text{pH}$   $6.9$ . The chemical composition of electrolytes in the injected water obtained for this study is given in Table 3.



**Table 3.** Chemical composition of *Rhoude El Baguel* injected water

Dosing elements		Concentration (mg/L)	Concentration (mEq)	
Cations	Ca <sup>2+</sup>	583.665	29.183	
	Mg <sup>2+</sup>	112.48	9.25	
	Fe <sup>2+</sup>	14.632	0.524	
	Na <sup>+</sup>	318.30	13.82	
	K <sup>+</sup>	25.98	0.66	
Total		-	53.437	
Anions	Cl <sup>-</sup>	1329.63	37.5	
	HCO <sub>3</sub> <sup>2-</sup>	169.58	2.78	
	SO <sub>4</sub> <sup>2-</sup>	574.48	11.96	
Total		-	52.24	
Salinity	1.6 mg/L	3.3 mS/Cm	pH	6.9

It should be noted that there is an influence of injection water's composition on SRB growth. Several authors [15-17] have mentioned the influence of physicochemical parameters on bacterial development. They stated that the growth of bacteria depends on the chemical composition of the medium, the physicochemical conditions and the specific nutrients present. Therefore the injection water studied contains nutrients (carbon, sulfur, and trace elements), which are essential to bacteria growth. Determination of mineral salts shows that injection water is salty and contains high concentration of sulfate ions, which play the role of electron acceptor [18]. In fact, sulfates also have a source of sulfur at SRB and are included in composition of two amino acids, cysteine and methionine which participate in proteins structure [19].

The presence of considerable concentration of Ca<sup>2+</sup> and Mg<sup>2+</sup> ions is observed; certainly, Ca<sup>2+</sup> ions favor bacterial absorption. On the other hand, Mg<sup>2+</sup> ions are in favor of reducing sulfates to sulfides [19]. We note that this water has a very low concentration of iron; in fact, this present -concentration comes from metal dissolution. In this regard, this element is considered an enzymatic factor [20]. Moreover, iron is part of many cytochromes structure; it also enters into basic structure of hydrogenases, the important enzymes of sulfate reduction by SRB. It should be noted that reduction of sulfate results in hydrogen sulfide (H<sub>2</sub>S) formation, which reacts with ferrous ions (Fe<sup>2+</sup>) of culture medium to obtain a black precipitate of iron sulfide (FeS). In our study, waters present a wide sulfate concentration. The SRB growth and their biological activity depend on sulfate concentration and the sampling water origin. In general, SRB tend to synthesize one or more metabolic products. Biochemically, SRB transform a sulfate to sulfide, a reaction which is catalyzed by adenosine phosphor-sulfate reductase.

### 3.2. Efficiency evaluation of bionatural extracts by microbiological method

In order to monitoring the efficiency of A, B, and C extracts on SRB growth as a function of incubation time, they were maintained at 37°C and over incubation period of 28 days. Before injection extracted products, a value of 10<sup>2</sup> germs/mL was obtained at the end of 14<sup>th</sup> day of incubation at 37°C. After injection 10 ppm of extract A, the concentration was zero until the 7<sup>th</sup> day of incubation, from there SRB concentration becomes 10 germs/mL up to the 28<sup>th</sup> day.

For the extract B injected at 10 ppm, the contamination does not remain during the entire incubation period. So, by injecting 10 ppm of extract C; no contamination while 14 days and from the 15<sup>th</sup> day it becomes equal to 10 germs/mL.

However, by injecting 20 ppm of each extract into penicillin vials containing 9mL of SRB contaminated injection water, extract A gave a zero concentration of bacterial germs for 14 days after which this concentration increased to 10 germs/mL. With extract B, no contamination was recorded during 28 days of incubation. Moreover, by injecting extract C, no contamination was observed for 21 days but at the end of the 22<sup>nd</sup> day a concentration of 10 germs/mL of SRB was noted. Of these results,

we find that the extract B obtained from *Gymnogongrus crenulatus* is very active against SRB growth. The extract C from *Pterocladia capillacea* is active at 10 ppm. Extract A from *Corallina elongata* is active at 20 ppm. To confirm these results, a study of efficiency by electrochemical techniques for three extracts A, B and C was carried out.

### 3.3. Efficiency evaluation of bionatural extracts on biocorrosion by metal weight loss method

After a 45-day contact between (the steel-injection water) which underwent treatment with different concentrations of plant extracts, we obtained the results given in Table 4.

**Table 4.** Evaluation of corrosion rate by weight loss method as a function of the variation in the concentration of the algae extract in the injection water

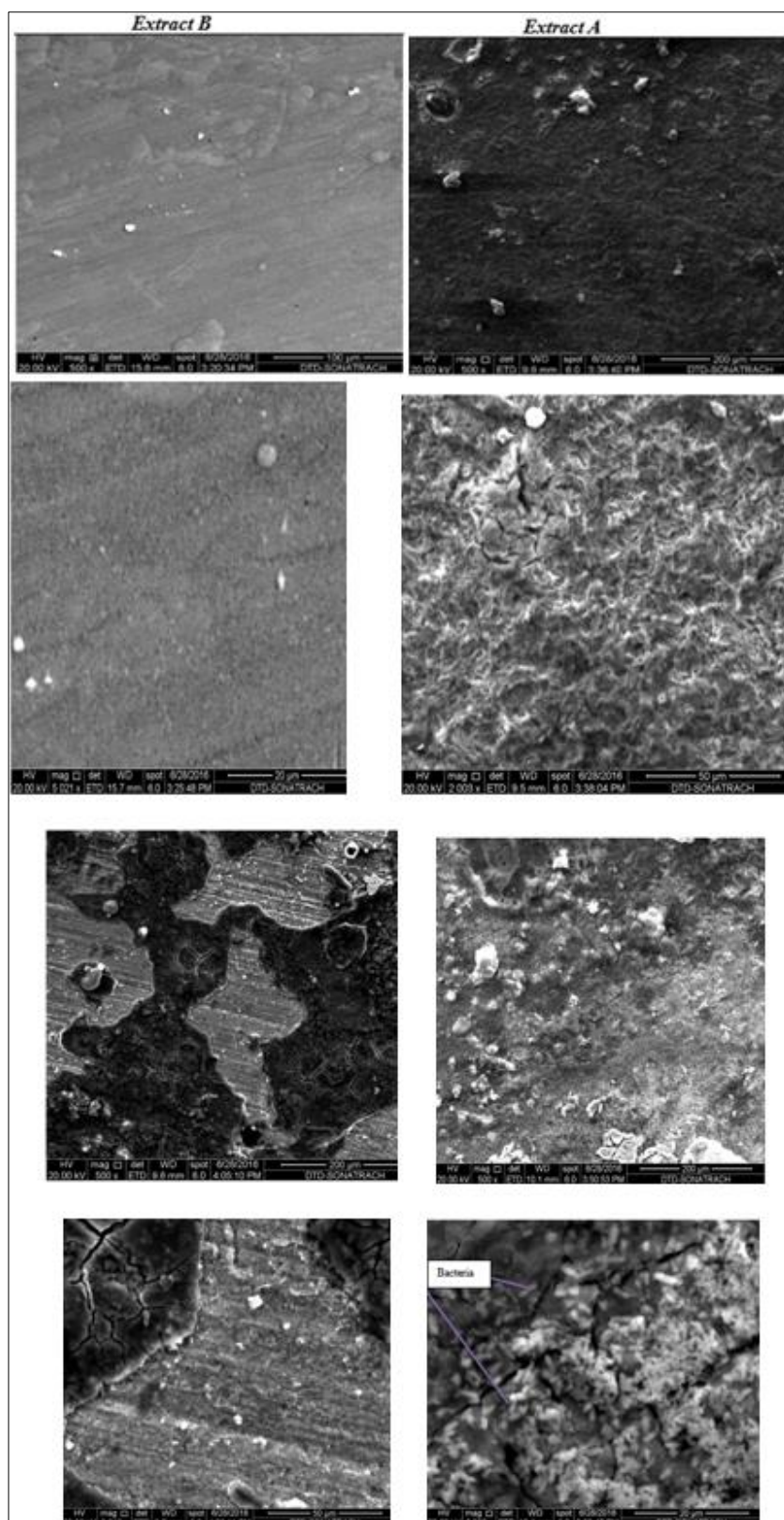
Carbon Steel API5LX60	Initial Weight $W_i$ (g)	Final weight $W_f$ (g)	$\Delta W$ (g) ( $W_i - W_f$ )	Exposure Time (Days)	Corrosion rate CR mm/year	Efficiency (%)
Control sample CS	8.68	4.9	<b>3.78</b>	45	<b>0.003</b>	-
extractA	9.64	9.58	<b>0.06</b>		<b>0.00004</b>	98.66
extractB	9.24	9.22	<b>0.02</b>		<b>0.00001</b>	99.66
extractC	8.39	8.36	<b>0.03</b>		<b>0.00002</b>	99.33

$$CR \text{ (mm/year)} = \Delta W / (\rho * \text{Time} * A) * 0.365; (\rho = 7.8 \text{ g/cm}^3); A: \text{Coupon area cm}^2$$

From results of Table 4, we find that corrosion rate of carbon steel in presence of SRB over a period of 45 days is high; it is 0.003 mm/year. After injection of the extract A, corrosion rate decreases and gives a protective power of 98.66 %. With extract B, corrosion rate is 0.00001 mm/year, which gives a protective power of 99.66 %. Finally, with the injection of extract C, corrosion rate is 0.00002 mm/year giving a protective power of 99.33%. We conclude that protective power of bionatural extract B is higher than C and higher than A. The active substance extracted from red algae *Gymnogongrus crenulatus* has a bactericidal power over corrosion caused by SRB more active than bionatural extract from *Corallina elongata* and *Pterocladia capillacea*.

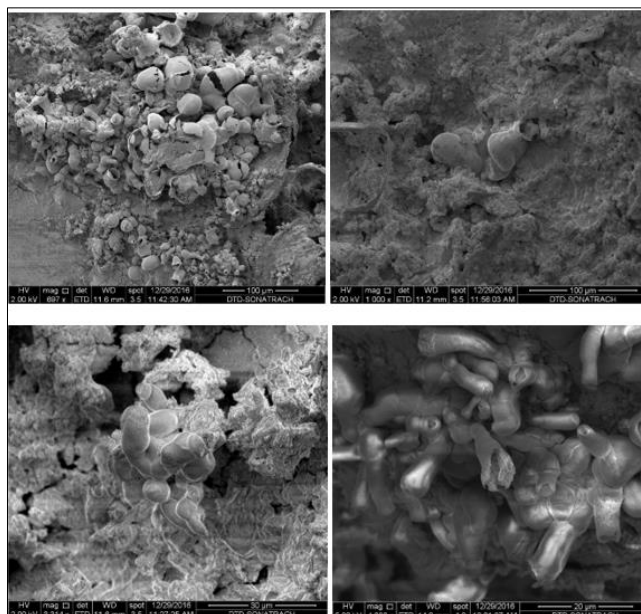
### 3.4. Surface morphology analysis

A microscopy analysis of morphology was performed using the scanning electron microscopy (SEM) and metallurgical characterization of the API 5L X60 carbon steel. Figures 4-7 show the surface after formation of the bacterial biofilm with and without treatment by the natural extracts A, B and C obtained from the red algae to distinguish the formed biofilm and the protective film of the biomolecules inhibiting biocorrosion. Metallurgical observations of carbon steel coupons immersed for 45 days in contaminated injection water by SRB and treated with bionaturels extracts A, B and C show differences between treatments performed surfaces.

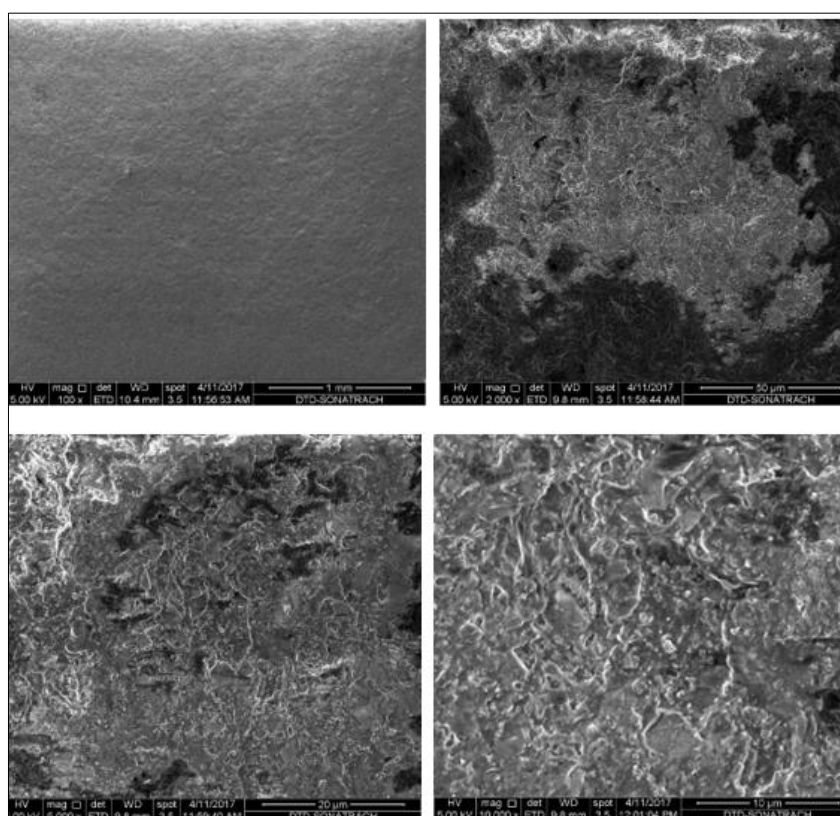


**Figure 4.** SEM images API 5L X60 carbon steel surface immersed for 45 days in injection water contaminated with SRB and treated with extracts B of 15 ppm concentration

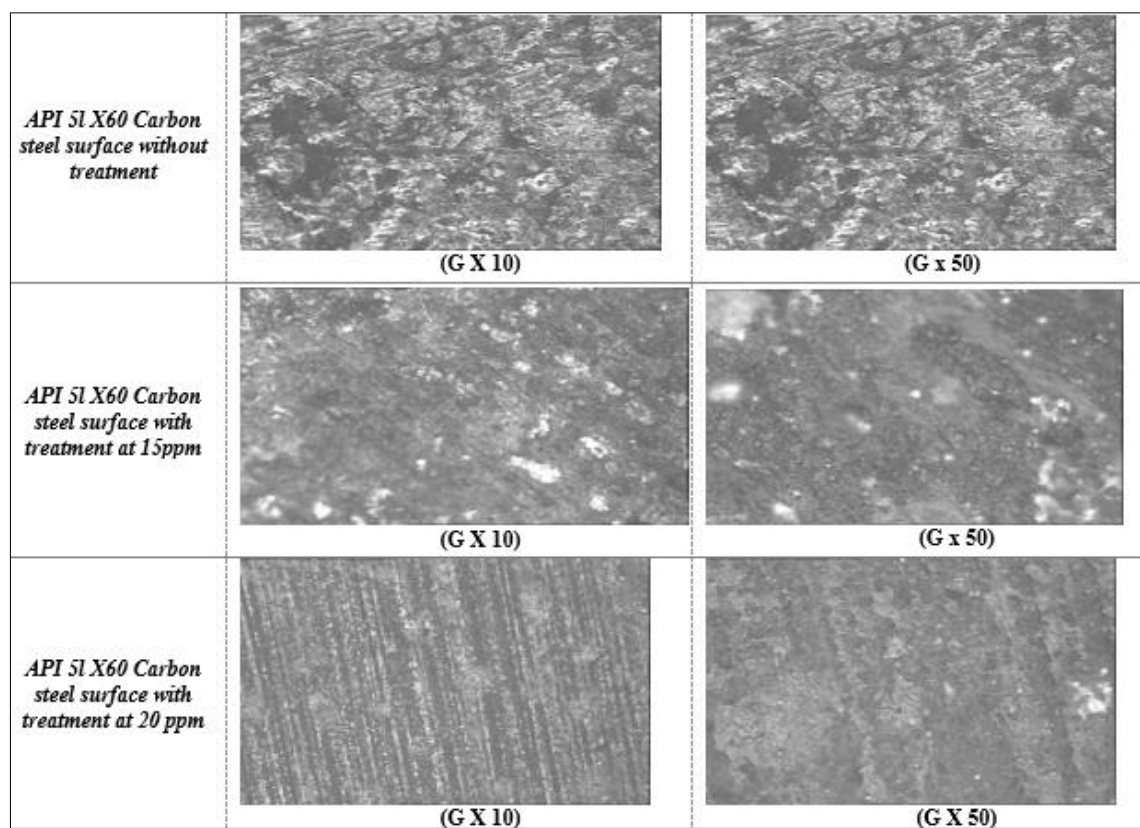




**Figure 5.** SEM Characterization by different magnifications of the carbon steel surface of API 5L X60 grade. Stage of formation of a bacterial Biofilm after immersion in injection water inoculated by SRB after 45 days of incubation at 37°C



**Figure 6.** SEM Characterization of carbon steel surface of API 5L X60 grade at different magnifications - Step of formation of a protective film after immersion in a contaminated injection water by the SRB and treated with the extract a obtained from *Corallina elongata* red algae extract after 45 days of incubation at 37°C

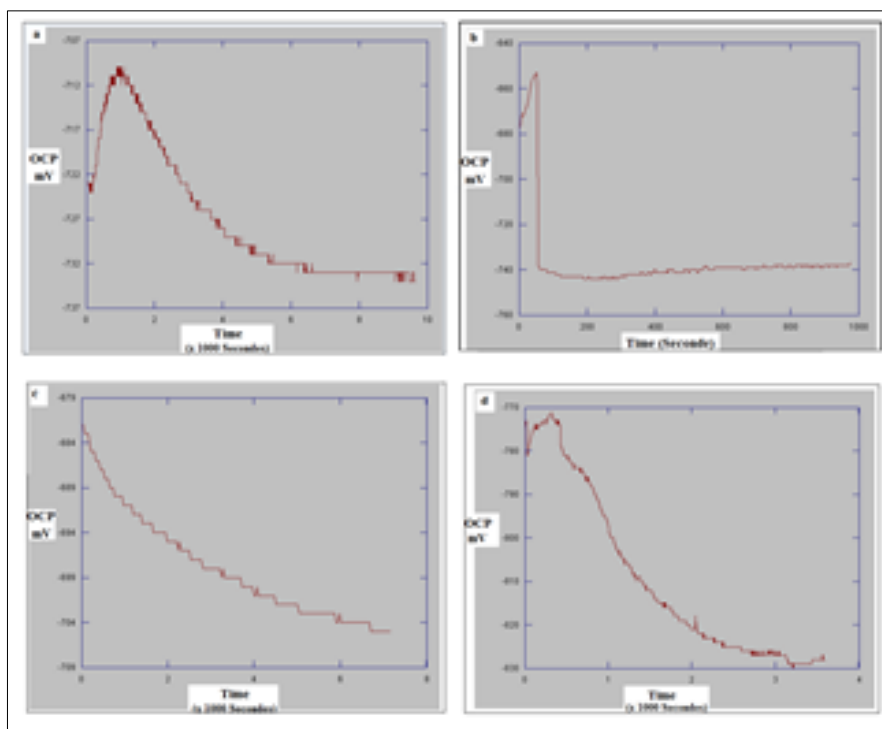


**Figure 7.** Metallurgical microscopy images of the coupons API 5L X60 Carbon steel surface without and with treatment after stripping the biofilm

The observation of the steel coupons with the scanning electron and metallurgical microscope shows more details. According to Figures 4-7, we noticed the presence of a biofilm formed by the sulfato-reducing bacterial consortium that covered the steel surface by the products of the bacterial metabolism formed during 45 days of immersion. Water is a changing environment where there is a trade-off between contaminated water and steel, bacterial biofilm deposit is formed on the metal surface. According to micrographics photos, SRB tend to focus on metal surfaces and associate in communities to form a biofilm. Thus, we shown that metal supports are susceptible to microbial colonization. At a concentration 15ppm and 20 ppm, the surface of the steel treated with the algae extract B is clear and completely clean, forms a protective layer on the metal, and prevents the development of bacterial biofilm. Extract B is more effective against corrosion by inhibiting the development of SRB.

### 3.5. Efficiency evaluation of bionatural extracts on biocorrosion by electrochemical techniques

Open-circuit potential (OCP) evolution over the time is of crucial importance. Certainly, the profile of the  $OCP = f(t)$  curve gives information on the evolution of the metal surface in contact with corrosive medium. The curves of potentials obtained as a function of time for the electrodes immersed in a solution contaminated by SRB before and after treatment with bionatural extracts A, B and C are shown in Figure 8. Table 5 presents the obtained data of OCP indicating stabilization time and incubation time.



**Figure 8.** Open-circuit potential evolution in the first three hours of carbon steel immersed in injection water treated with bionatural extracts A, B and C – **a.** Control sample – **b.** Treat by 10 ppm of extract A after 5 days immersion – **c.** Treated by 10 ppm of extract C after 12 days immersion – **d.** Treated by 10 ppm of extract B after 10 days immersion

**Table 5.** Open- circuit orrosion potential of carbon steel measured in injection water contaminated by SRB and treated by bionatural extracts A, B and C

Injection water inoculated with SRB	Incubation Time (Days)	Stabilisation time (Second)	OCP V/SCE
without treatment	3	9600	-0.735
treated with 10 ppm of extract A	5	3600	-0.698
treated with 50 ppm of extract A.	6	3700	-0.663
treated with 10 ppm of extract B	10	3600	-0.677
treated with 50 ppm of extract B	11	3900	-0.533
treated with 10 ppm of extract C	12	3600	-0.671
treated with 50 ppm of extract C	13	3900	-0.633

OCP evolution gives stabilization times that differ from one electrolyte to another according to the species present in each medium (injection water, sulfate-reducing bacteria, extract A, extract B, extract C). Without treatment, OCP tends towards more negative values (-0.735 V/SCE), indicating presence of corrosion in this area [21]. By injecting different concentrations from 10 ppm to 50 ppm of bionaturels extracts, OCP with extract A gives -0.698 and -0.663 V/SCE respectively. Indeed, OCP with extract B gives -0.677 and -0.533 V/SCE and with extract C gives -0.671 and -0.633 V/SCE. These values tend

towards a more noble or electropositive potential. So, we are in protection domain according to the Pourbaix diagram [21]. After potential stabilization, the system allows us to test other electrochemical techniques such as LRP and EIS test to determine the phenomena occurring at the metal/solution interface. Concerning, linear polarization resistance measurements using Resistance the bionatural extract A, the values of polarization resistance  $R_p$  obtained from the slopes of straight lines electrode potential =  $f$  (current density) for the system (injection water and chemical species of extract A from red algae *Corallina elongata* at different concentrations) in contact with carbon steel are reported in Table 6. Values of corrosion current density, corrosion rate and inhibitory efficiency are also listed.

**Table 6.** Electrochemical parameters obtained by LRP technique of carbon steel immersed in an untreated medium and treated one with extract A

Concentration of bionatural extract A (ppm)	OCP (V/SCE)	Polarization resistance $R_p$ ( $K \Omega \cdot cm^2$ )	Corrosion current density $I_{Corr}$ ( $\mu A/cm^2$ )	CR (mm/year)	Efficiency (%)
0	-0.735	0.205	105.48	2.088	/
5	-0.698	1.485	14.62	0.148	86.20
10	-0.698	2.112	10.27	0.104	90.29
15	-0.695	3.125	06.94	0.070	93.44
20	-0.697	4.205	05.09	0.051	95.12
30	-0.682	6.115	04.79	0.048	96.65
50	-0.663	7.338	02.95	0.033	97.20

From the results reported in Table 6, we find low strength of steel without treatment. We noted the value of  $0.205 K\Omega \cdot cm^2$  that corresponds to 2.088 mm/year corrosion rate. After injection of different concentrations of bionatural extract A, results analysis shows that carbon steel resistance polarization as well as extract efficiency increase proportionally with increase in concentration of extract injected to achieve  $1.485 K\Omega \cdot cm^2$  and an efficiency of 86.20% at 5 ppm. By injecting 30 ppm of extract, corrosion rate is 0.048 mm/year was recorded and the processing efficiency reached 96.65%.

Concerning the bionatural extract B, the values of polarization resistance obtained from results exploitation of current intensities  $OCP = f(I)$  of system (injection water and chemical species of bionatural extract B extracted from red algae *Gymnogongrus crenulatus* at different concentrations in contact with carbon steel API5LX60 are reported in the Table 7.

**Table 7.** Electrochemical parameters obtained by LRP technique of carbon steel immersed in an untreated medium and treated one with bionatural extract B

Concentration of bionatural extract B (ppm)	OCP (V/ECS)	LRP ( $K \Omega \cdot cm^2$ )	$I_{Corr}$ ( $\mu A/cm^2$ )	CR (mm/year)	Efficiency (%)
0	-0.735	0.205	105.481	2.088	/
5	-0.660	67.230	0.322	0.0064	99.69
10	-0.677	67.365	0.321	0.0064	99.69
15	-0.667	67.500	0.343	0.006	99.69
20	-0.677	68.850	0.343	0.006	99.70
30	-0.670	71.830	0.302	0.0059	99.71
50	-0.533	73.899	0.293	0.0058	99.72

From results in Table 7, it was found that injection of bionatural extract B gave an excellent efficiency as its concentration in solution contaminated by SRB increased, at 5ppm efficiency exceeds 99% (99.69%) and at 30 ppm the efficiency increases again to reach 99.71%. These values are followed by high polarization resistances of carbon steel and by low corrosion rates (0.005 mm/year). Low corrosion current densities  $I_{Corr}$  are recorded. this may be justified by the fact that the free bioactive molecules in the solution tend to adsorb on steel surface, consequently, the increase in covering of protective film on



surface of steel against biocorrosion prevents the passage of current which activates oxidation-reduction reactions, hence the blocking of corrosion phenomenon in general and of biocorrosion in particular.

So, the bionatural extract C, the values of polarization resistance obtained from results exploitation of current intensities  $OCP = f(I)$  of system (injection water and chemical species of bionatural extract C obtained from the red alga *Pterocladia capillacea* at different concentrations in contact with carbon steel API5LX60 are reported in the Table 8. It can be seen that polarization resistance of carbon steel immersed in injection water contaminated with SRB is low; it is  $0.205 \text{ K}\Omega\cdot\text{cm}^2$  that corresponds to  $2.088 \text{ mm/year}$ . After injection bionatural extract C at different concentrations, an increase in polarization resistance of steel was recorded as well as inhibitory efficiency which is proportional with increase of concentration to reach a resistance of  $7.801 \text{ k}\Omega\cdot\text{cm}^2$  And an efficiency of  $97.37\%$  at  $5\text{ppm}$ . That confirms protective layer formation on steel surface. By increasing the concentration of extract, protective film becomes more important this is confirmed by recording of the values of resistances of polarization which reaches  $22.917 \text{ K}\Omega\cdot\text{cm}^2$  and a protective efficiency of  $99.10\%$  (Table 8).

**Table 8.** Electrochemical parameters obtained by lrp technique of carbon steel immersed in an untreated medium and treated one with bionatural Extract C

Concentration of bionatural extract C (ppm)	OCP (V/ECS)	LRP ( $\text{K}\Omega\cdot\text{cm}^2$ )	$I_{\text{corr}}$ ( $\mu\text{A}/\text{cm}^2$ )	CR (mm / year)	Efficiency (%)
0	-0.735	0.205	105.185	2.088	-
5	-0.677	7.801	2.782	0.055	97.37
10	-0.671	9.360	2.319	0.032	97.80
15	-0.654	13.482	1.610	0.031	98.48
20	-0.656	15.822	1.372	0.027	98.70
30	-0.644	20.128	1.053	0.026	98.98
50	-0.633	22.917	0.824	0.018	99.10

Regarding the use of electrochemical impedance spectroscopy (EIS), this technique showed that when carbon steel is immersed in water inoculated with SRB, the metal surface is covered with a biofilm of microorganisms. Thus, the biofilm formed affects the transfer of charge and alters corrosion process of steel. EIS measurements have been carried out to monitor its formation and its evolution. This technique makes it possible to dissociate different steps of interfacial process, by separate evidencing values of ohmic resistance of electrolyte ( $R_e$ ) from charge transfer resistance ( $R_{ct}$ ), it also makes possible to calculate differential capacitances of electrical double layer ( $C_{dl}$ ) [22]. The results are generally represented by Nyquist and Bode diagrams, the response of an interface formed diameter of second semicircle is  $R_{ct}$ . The first characteristic is biofilm and second characterizes corrosion processes of metal [23].

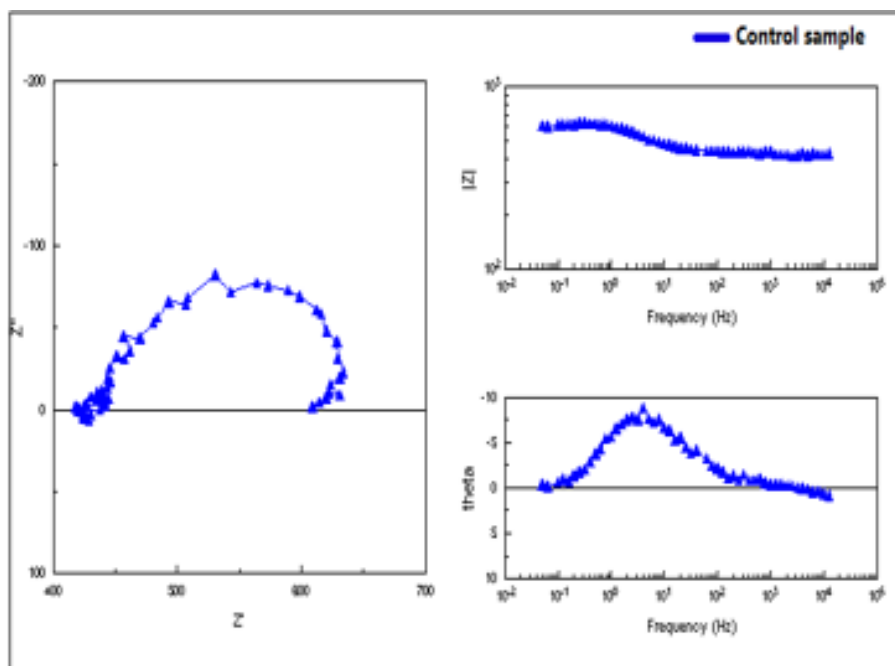
When we used the extract A and based on Figure 9, we found out that semicircle impedance diagram characterizes carbon steel corrosion control by charge transfer process.  $R_e$  was low for solution inoculated by SRB and not treated by bionatural extract A, according to impedance parameters given in Table 9, an increase in  $R_{ct}$  and a decrease in electrical  $C_{dl}$  values as a function of increase in concentration of bionatural treatment extract. These results allow us to conclude that extract A inhibits process of carbon steel biocorrosion by adsorption mechanism. It is also noted that  $C_{dl}$  is inversely proportional to  $R_p$  calculated previously. Higher resistance of material, the greater formation of protective film on metal surface, and more electronic ion-exchanging cloud (anions, cations).  $C_{dl}$  was between  $10^{-5}$  and  $10^{-6}$  Farad/ $\text{cm}^2$ . The biomolecule inhibitory action by adsorption mechanism on metal surface is advanced. Certainly, more extract A was adsorbed to the surface, capacity of double layer was lower.

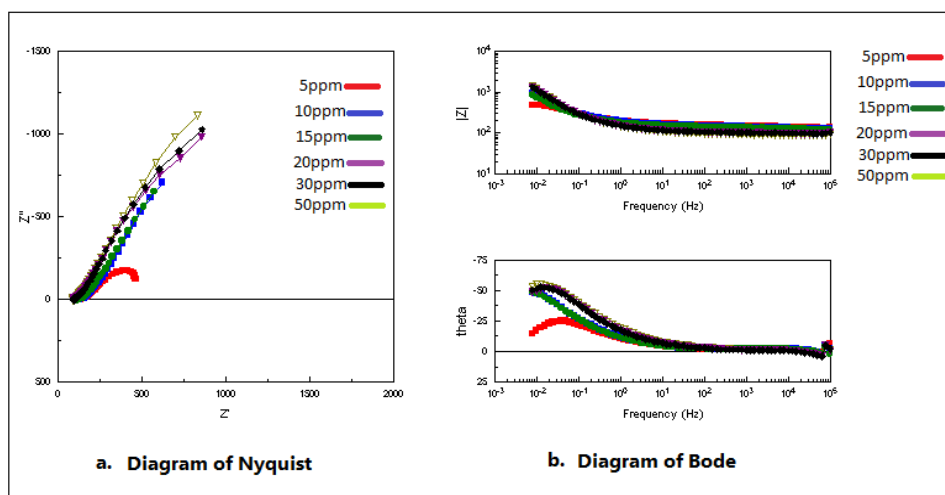


When we used the extract B and based on Figure 10 and from values of impedance parameters given in Table 10, we noted an increase in  $R_{tc}$  and a decrease in  $C_{dl}$  values as a function of the increase in concentration of bionatural extract B. These results allow us to conclude that biocorrosion inhibition process is associated with the adsorption of molecules of algae extract on steel surface indicated by phase angle  $\Theta$ . The maximum phase angle values were is  $-40^\circ$  and  $-42^\circ$  at 5 ppm and at 20 ppm concentrations respectively, with respect to the doses of 10 ppm, 15ppm, 30ppm and 50ppm the scanning of material surface was performed by two angles phase, which confirms presence of protective film of bionatural extract B on steel surface.

**Table 9.** Electrochemical parameters obtained by eis technique of carbon

Concentrations (ppm) Parameters	0	5	10	15	20	30	50
OCP (V/ECS)	-0.735	-0.749	-0.754	-0.759	-0.660	-0.672	-0.675
$R_s$ (k. $\Omega$ Cm <sup>2</sup> )	0.02	0.13	0.09	0.09	0.12	0.12	0.04
$R_{tc}$ (k. $\Omega$ Cm <sup>2</sup> )	0.25	1.04	1.85	2.10	3.52	5.44	7.20
$R_p$ (k. $\Omega$ Cm <sup>2</sup> )	0.27	1.17	1.94	2.19	3.64	5.56	7.24
$C_{dl}$ ( $\mu$ F/Cm <sup>2</sup> )	200	40.50	40.50	2.92	2.98	2.98	1100
Maximum phase Angle $\Theta$ (deg)	$-8^\circ$	$-24^\circ$	$-51^\circ$	$-53^\circ$	$-48^\circ$	$-48^\circ$	$-44^\circ$
Efficiency (%)	/	76.877	86.06	87.60	92.55	95.13	96.26

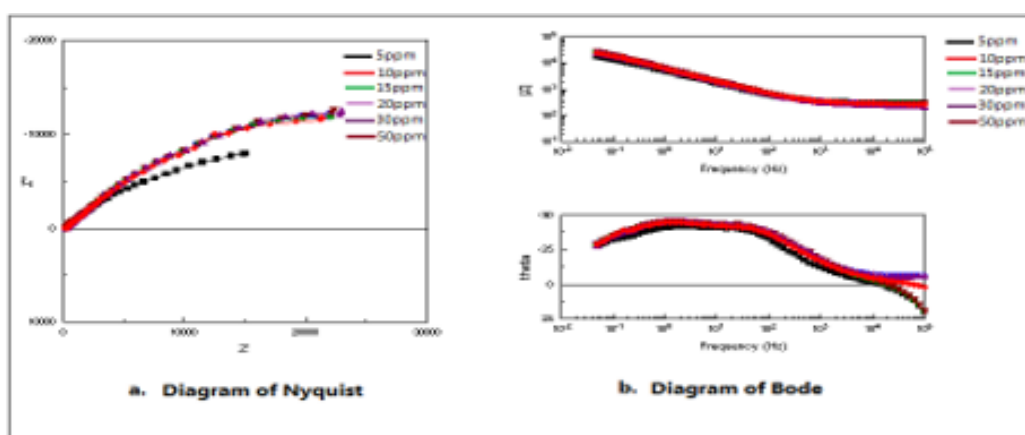




**Figure 9.** Nyquist diagram and Bode diagram of carbon steel immersed in injection water contaminated with SRB, treated and untreated by bionatural extract A

**Table 10.** Electrochemical parameters obtained by eis technique of carbon steel immersed in an untreated medium and treated one with bionatural extract B at different concentrations

Concentrations ppm Parameters	0	5	10	15	20	30	50
(V/ECS)	-0.735	-0.830	-0.667	-0.649	-0.656	-0.660	-0.649
$R_s$ ( $k \Omega \cdot \text{cm}^2$ )	0.02	0.29	0.27	0.26	0.27	0.26	0.23
$R_{ct}$ ( $k \Omega \cdot \text{cm}^2$ )	0.25	40.25	50.66	51.25	51.33	52.42	53.15
$R_p$ ( $k \Omega \cdot \text{cm}^2$ )	0.27	40.55	50.93	51.51	51.60	52.68	53.37
$C_{dl}$ ( $\mu\text{F}/\text{cm}^2$ )	210	140	79	76.8	74.2	80.5	71.5
Maximum phase Angle $\theta$ (deg)	$-8^\circ$	$-40^\circ$	$-41^\circ; -44^\circ$	$-44^\circ; -41^\circ$	$-42^\circ$	$-44^\circ; -42^\circ$	$-45^\circ; -42^\circ$
Efficiency (%)	/	99.33	99.46	99.47	99.47	99.48	99.49



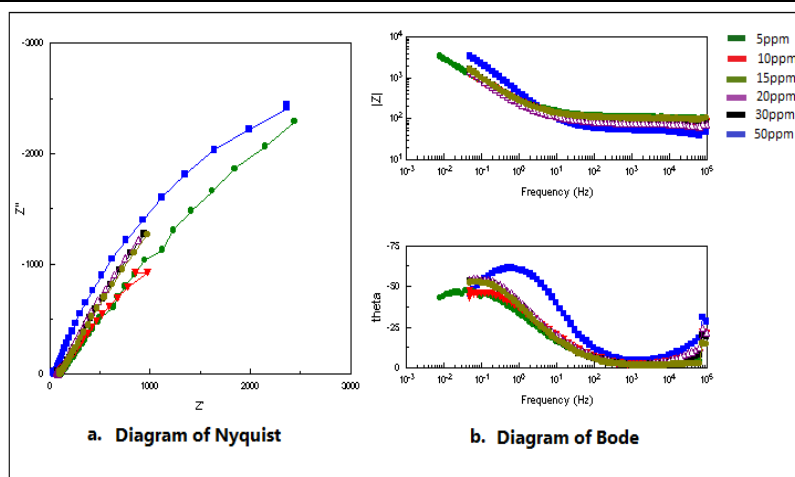
**Figure 10.** Nyquist diagram and Bode diagram of carbon steel immersed in injection water contaminated with SRB, treated and untreated by bionatural extract B

Concerning the bionatural extract C, based on impedance parameters given in Table 11 and shown on Figure 11, we noted an increase in  $R_{ct}$  and a decrease in  $C_{dl}$  values as a function of increase in concentration of bionatural extract C. These results allow us to announce that our algae extract inhibits biocorrosion process of carbon steel by adsorption mechanism.  $C_{dl}$  is inversely proportional to  $R_p$ .

Higher resistance of material gives as, the more  $C_{dl}$  decreases until values of  $10^{-5}$  Farad/cm<sup>2</sup> are obtained. Inhibitory action of biomolecule by adsorption mechanism on metal surface was confirmed.

**Table 11.** Electrochemical parameters obtained by eis technique of carbon steel immersed in an untreated medium and treated one with bionatural extract c at different concentrations

Concentrations ppm Parameters	0	5	10	15	20	30	50
OCP (V/ECS)	-0.735	-0.961	-0.958	-0.936	-0.956	-0.937	-0.941
$R_s$ (k $\Omega$ . Cm <sup>2</sup> )	0.02	0.09	0.10	0.04	0.10	0.10	0.10
$R_{tc}$ (k $\Omega$ . Cm <sup>2</sup> )	0.25	5.47	6.10	10.39	11.01	14.14	14.31
$R_p$ (k $\Omega$ . Cm <sup>2</sup> )	0.27	5.56	6.20	10.43	11.11	14.24	14.41
$C_{dl}$ ( $\mu$ F/Cm <sup>2</sup> )	210	4100	800	300	300	300	26
Maximum phase Angle $\theta$ (deg)	-8°	-38°	-46°	-63°	-52°	-50°	-48°
Efficiency (%)	/	95.13	95.63	97.40	97.56	98.09	98.12



**Figure 11.** Nyquist diagram and Bode diagram of carbon steel immersed in injection water contaminated with SRB, treated and untreated by bionatural extract C

## 4. Conclusions

We studied inhibition efficiency treatment of biocorrosion with natural extracts from three species of marine red algae that we identified: *Corallina elongata*, *Gymnogongrus crenulatus* and *Pterocladia capillacea*. The ethanol extracts were tested as corrosion inhibitors on carbon steel immersed in injection water contaminated with sulfato-reducing bacteria (SRB). Infrared spectra of three extracts from red algae confirmed the presence of molecules, which contain amine functional groups (trimethylamines) that act as inhibitors of bacterial corrosion by their protective properties. The physicochemical analyzes of injection water showed that it is rich in nutriment such as carbon, sulfur, magnesium that are essential for bacterial development. The microbiological analysis of this water revealed  $10^2$  germs/mL of SRB concentration. Corrosion rate was firstly determined by weight loss method. The data have shown that the active substance extracted from red algae *Gymnogongrus crenulatus* has a higher bactericidal power over corrosion caused by SRB, more active than the extracts from *Corallina elongata* and *Pterocladia capillacea*. SEM and metallographic observations have confirmed the presence of protective layer formed on steel surface.

The use of electrochemical techniques (OCP-time, LPR and EIS) permitted to determine various electrochemical parameters (corrosion current density, corrosion rate, polarization resistance, inhibition efficiency, electrolyte resistance, charge transfer resistance, double layer capacity of protective film), these results making possible to evaluate protection properties of bionatural extracts. The shift of

corrosion potential to electropositive values with increased extract concentration in electrolyte indicates the inhibition ability of each extract. Electrochemical impedance diagrams showed Nyquist semicircles which increase their diameter proportionally with extract concentration. Similarly, the values of charge transfer  $R_{ct}$  were comparable to  $R_p$  obtained from LRP techniques. Thus, both resistances represent the ability of steel surface to prevent flow of current into the medium. The electrochemical data showed an increase in protective power and a decrease in corrosion rate as extract concentration increased. The extract from *Gymnogongrus crenulatus* has given a very effective protection to stop carbon steel corrosion. By comparing our results with previous works we deduced that bionaturels algae extracts have a good anticorrosive effect and can have their place in the petroleum industry.

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